

# Fibrinogen Disorders Panel

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**Versiti offers comprehensive genetic analysis to detect sequence variants and large deletions and duplications in 3 genes known to cause fibrinogen disorders. This panel can be ordered as:**

- **Next Generation Sequencing (NGS) only;**
- **NGS with reflex to Array Comparative Genomic Hybridization (aCGH) Deletion/Duplication if sequencing does not identify clinically significant variants that fully explain the clinical or laboratory phenotype;**
- **NGS analysis with concurrent aCGH Deletion/Duplication (both testing methodologies performed simultaneously); or**
- **Deletion/Duplication by aCGH only.**

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Inherited fibrinogen disorders are a group hemostatic disorders characterized by bleeding and/or thrombosis due to quantitative (afibrinogenemia, hypofibrinogenemia) and/or qualitative defects (dysfibrinogenemia, hypodysfibrinogenemia) in fibrinogen. They are inherited in an autosomal dominant or autosomal recessive pattern. Fibrinogen is a hexamer of three subunits (alpha, beta and gamma) encoded by the *FGA*, *FGB* and *FGG* genes respectively. Fibrinogen is converted to fibrin by thrombin to form a fibrin-based clot. Fibrin is also a negative regulator of thrombin and participates in the cellular processes leading to revascularization and wound healing.

Inherited fibrinogen disorders are rare, with an estimated incidence of 1 in one million births. Patients present with a wide range of clinical manifestations and severity; from severe, lifelong, delayed bleeding (including bleeding and

impaired healing of the umbilical stump) to minimal/absent bleeding and/or thrombosis. A prolonged thrombin time, proportionally decreased fibrinogen antigen and activity, or significantly decreased and discrepant activity compared to antigen is observed on laboratory testing.

Afibrinogenemia (severe quantitative defect) is usually caused by homozygosity (and less frequently compound heterozygosity) of pathogenic null variants, while hypofibrinogenemia, dysfibrinogenemia and hypodysfibrinogenemia are caused by heterozygous pathogenic variants, the majority of which are missense, although nonsense, frameshift and splice site variants have also been described.

The NGS panel evaluates for single nucleotide variants and small deletions and duplications, which are most commonly responsible for genetic disease. However, large deletions and duplications, also referred to as copy number variations (CNVs), are a known cause of genetic disorders, but can escape detection by next generation sequence analysis. Additional testing with aCGH Deletion/Duplication analysis is available for all genes on this panel to evaluate for large deletions and duplications encompassing one or more exons, or affecting an entire gene.

Refer to the table for further information about each gene in the Fibrinogen Disorders Panel, including the clinical phenotype and inheritance pattern.

## Fibrinogen Disorders Panel: gene, clinical phenotype, population-specific comments and inheritance pattern

Gene	Clinical Phenotype	Inheritance
FGA FGB	Afibrinogenemia: severe/delayed bleeding from markedly decreased or absent fibrinogen	Autosomal Recessive
	Hypofibrinogenemia: mild to moderate delayed bleeding due to decreased fibrinogen levels	Autosomal Dominant (most common)/ Autosomal Recessive
FGG	Hypodysfibrinogenemia: mild to moderate delayed bleeding with or without thrombosis due to deficient and dysfunctional fibrinogen	Autosomal Dominant (most common)/ Autosomal Recessive
	Dysfibrinogenemia: absent or mild/moderate delayed bleeding with or without thrombosis due to dysfunctional fibrinogen	Autosomal Dominant (most common)/ Autosomal Recessive

### Indications for testing:

#### Fibrinogen Disorders Panel (NGS and/or aCGH), order code 4885:

The Fibrinogen Disorders Panel should be considered:

- In patients presenting with bleeding and/or thrombosis and decreased fibrinogen activity or discrepancy between fibrinogen antigen and activity, especially when suspected to be inherited
- In patients with familial venous thrombosis, where no defect was identified after testing for other more common causes of inherited thrombophilia
- To clarify possible carrier status in individuals who have a family history of a fibrinogen disorder, but the causal familial variant(s) is unknown.

#### Single Gene Analysis (order code 4855) or Custom Blood Disorder Panel (Order Code 4850), (NGS and/or aCGH):

Analysis of genes included in this panel may also be ordered as standalone Single Gene Analysis or as a Custom Blood Disorder Panel (2-10 genes), by NGS and/or by aCGH, as dictated by the patient's clinical and laboratory phenotype, as well as their ancestry, or to supplement previous genetic testing.

#### Targeted Familial Variant Analysis (order code 4970):

Targeted variant analysis for clinical diagnosis, carrier identification, or prenatal diagnosis can also be performed on any gene in the panel when the pathogenic variant(s) is known in the family. If the proband was not tested at Versiti, a control sample is preferred and may be required (please call the laboratory to discuss). If the familial variant is a large deletion or duplication, aCGH for the involved gene is required.

For clinical questions about laboratory tests and test utilization support, contact Versiti Client Services: (414) 937-6396 or 800-245-3117, Option 1, to be directed to our genetic counselors and clinical support team.

### Test method:

**NGS:** This next-generation sequencing assay analyzes the complete coding region of 3 genes plus a minimum 30bp of non-coding DNA, including intron-exon boundaries, and is compared to the build GRCh37.p13 reference sequence. These targeted regions are captured by hybridization, amplified, and sequenced by massively parallel sequencing. Regions will have a minimum coverage of 50x and those regions with less than 50 sequencing reads or low quality coverage are supplemented with Sanger sequencing. All regions are covered by bidirectional analysis. Variants are identified by a customized bioinformatics pipeline, analyzed and comprehensively interpreted by our team of practicing hematologists with expertise in non-malignant hematology and laboratory diagnostics, scientists, and genetic counselors. All reported variants, including pathogenic, likely pathogenic, and variants of uncertain significance, are confirmed by Sanger sequencing. For prenatal testing, analysis of variable number tandem repeats (VNTR) is used to confirm results are not affected by maternal cell contamination.

**aCGH:** The specific genes are analyzed for copy number variations due to deletion or duplication by high density gene-focused array Comparative Genomic Hybridization. Probes are approximately 60bp in length and density of coverage in exonic regions is a minimum of 4 probes per 500 bp. Genomic DNA for the samples and gender-matched references are denatured, labeled with fluorescent dye and hybridized, the array is washed and scanned, and analysis is performed for the specific genes requested.

### Assay sensitivity and limitations:

**NGS:** The analytical sensitivity of the NGS test is >99% for single nucleotide changes and insertions and deletions of less than 20 bp. NGS analysis is not designed to detect large deletions or duplications (>20 bp), or variants that are outside the regions sequenced. Low level mosaicism will not be detected by this sequencing methodology.

**aCGH:** Balanced chromosomal rearrangements (i.e., translocations, inversions) or point mutations that may be the cause of the clinical phenotype cannot be detected via aCGH. Any exonic deletion or duplication smaller than 500bp may not be detected. Low level of mosaicism will not be detected by aCGH. Probe performance could be affected by multiple SNPs in a given region. Breakpoints occurring outside the targeted gene(s) will not be defined.

### Clinical Sensitivity

The clinical sensitivity of comprehensive genetic testing (NGS and aCGH) of the 3 genes known to be associated with Fibrinogen Disorders is highest in patients presenting with bleeding and/or thrombosis with decreased fibrinogen activity or discrepancy between fibrinogen antigen and activity, that is suspected to be inherited.

### Reporting of Results

Results are classified and reported in accordance with ACMG next-generation sequencing and copy number variation standards. Sequence variants and large deletions and duplications predicted to be pathogenic, likely pathogenic, and of uncertain significance will be reported; variants classified as likely benign or benign are typically not reported but such data are available upon request. Sequence variants are described using standard Human Genome Variation Society (HGVS) nomenclature (<http://hgvs.org>); copy number variants are described in accordance with the International System for Human Cytogenomic Nomenclature (ISCN).

### Specimen Requirements

**Parental/Patient/Pediatric:** 3-5 mL Whole blood (EDTA tube, lavender top), 2-5 mL Bone marrow (EDTA tube, lavender top), 3-4 Buccal swabs, or  $\geq 1\mu\text{g}$  of DNA at  $\geq 50\text{ng}/\mu\text{L}$  of High Quality DNA.

**Fetal:** 7-15 mL amniotic fluid, 5-10 mg chorionic villi; back up culture of amniocytes or chorionic villi is highly recommended. Cultured: Two T25 flasks cultured amniocytes or chorionic villi ( $2 \times 10^6$  minimum). Maternal blood sample of 3-5 mL Whole blood (EDTA tube, lavender top) is requested for all prenatal samples for maternal cell contamination studies. For questions please contact the laboratory to discuss sample requirements.



SHIP

### Shipping Requirements

Ship on an ice pack or at room temperature. Protect from freezing. Place the specimen and the requisition into plastic bags and seal. Insert into a Styrofoam container, seal and place into a sturdy cardboard box, and tape securely. Ship the package in compliance with your overnight carrier guidelines. Label with the following address:

Client Services/Diagnostic Laboratory  
Versiti  
638 N. 18th St  
Milwaukee, WI, 53233



ORDER

### Required Forms

Please complete all pages of the requisition form. Clinical history (including patient's ethnicity, clinical diagnosis, family history, and relevant laboratory findings) is necessary for optimal interpretation of genetic test results and recommendations. Clinical and laboratory history can either be recorded on the

requisition form or clinical and laboratory reports can be submitted with the sample.

### CPT Codes/Billing/Turnaround Time

**Test code:** 4885

For suggested CPT codes, visit the [versiti.org/test-catalog](https://www.versiti.org/test-catalog)

**Turnaround time:** NGS only, aCGH only, or NGS and aCGH concurrently: 21 days

NGS reflex to aCGH: 21 days (if NGS only, aCGH not needed) or 42 days (with reflex to aCGH)

The CPT codes provided are subject to change as more information becomes available. CPT codes are provided only as guidance to assist clients with billing.

For additional information related to shipping, billing or pricing, please contact Versiti Client Services: (414) 937-6396 or 800-245-3117, Option 1, or [LabInfo@versiti.org](mailto:LabInfo@versiti.org)

## References

### Fibrinogen disorders references

1. Acharya SS, Dimichele DM. Rare inherited disorders of fibrinogen. *Haemophilia*. 2008;14(6):1151-1158. doi:10.1111/j.1365-2516.2008.01831.x
2. Casini A, Blondon M, Tintillier V, et al. Mutational Epidemiology of Congenital Fibrinogen Disorders. *Thromb Haemost*. 2018;118(11):1867-1874. doi:10.1055/s-0038-1673685
3. Jain S, Acharya SS. Inherited disorders of the fibrinolytic pathway. *Transfus Apher Sci*. 2019;58(5):572-577. doi:10.1016/j.transci.2019.08.007

### Variant interpretation references

4. Bean LJH, Funke B, Carlston CM, et al. Diagnostic gene sequencing panels: from design to report-a technical standard of the American College of Medical Genetics and Genomics (ACMG). *Genet Med*. 2020;22(3):453-461. doi:10.1038/s41436-019-0666-z
5. Rehm HL, Bale SJ, Bayrak-Toydemir P, et al. ACMG clinical laboratory standards for next-generation sequencing. *Genet Med*. 2013;15(9):733-747. doi:10.1038/gim.2013.92
6. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med*. 2015;17(5):405-424. doi:10.1038/gim.2015.30
7. Riggs ER, Andersen EF, Cherry AM, et al. Technical standards for the interpretation and reporting of constitutional copy-number variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics (ACMG) and the Clinical Genome Resource (ClinGen). *Genet Med*. 2020;22(2):245-257. doi:10.1038/s41436-019-0686-8

